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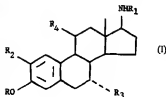
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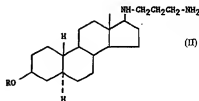
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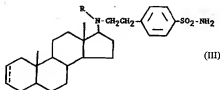
(54) Title: CYCLIC HYDROCARBONS WITH AN AMINOALKYL SIDECHAIN



(I)



(II)



(III)

(57) Abstract

This invention provides novel cyclic hydrocarbons of formula (I), wherein R is selected from the group consisting of $\text{CH}_2=\text{CH}-\text{CH}_2-$, $\text{HO}-\text{CH}_2\text{CH}_2\text{CH}_2-$, and CH_3 ; wherein R_1 is selected from the group consisting of *m*-trifluoromethylphenylmethyl, 2-thienylmethyl, and *p*-aminosulfonylphenylethyl; wherein R_2 and R_3 are methyl or hydrogen; wherein R_4 is hydrogen or -OH; a compound of formula (II), wherein R is $(\text{CH}_3)_2\text{NCH}_2\text{CH}_2\text{CH}_2-$ or $\text{NH}_2\text{CH}_2\text{CH}_2\text{CH}_2-$; and pharmacologically acceptable salts thereof, or a compound of formula (III), wherein the dashed line indicates that the 2-3 bond is saturated or unsaturated and, wherein R is hydrogen or methyl. These compounds are useful for inhibiting adverse physiological symptoms associated with phospholipase A_2 and for treating hyperglycemia associated diseases such as diabetes and obesity.

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CYCLIC HYDROCARBONS WITH AN AMINOALKYL SIDECHAINFIELD OF INVENTION

This invention encompasses novel cyclic hydrocarbons substituted with an aminoalkyl sidechain that are useful for inhibiting adverse
5 physiological symptoms associated with phospholipase A2 and for treating hypoglycemia associated diseases such as diabetes and obesity.

BACKGROUND

Phospholipase A2 hydrolyses arachidonic acid containing phospho-
10 lipids, thereby providing substrate to the multiple enzymes of the arachidonic acid cascade. The metabolites of the arachidonic acid cascade are varied and include prostaglandins, thromboxanes, leukotrienes, or other hydroxylated derivatives of arachidonic acid. The role of phospholipase A2 in the formation of prostaglandins in
15 mammalian metabolism is well known, see-W. Vogt, Advances in Prostaglandins and Thromboxane Research, 3, p. 89 (1978) and P.C. Isakson, et al., Advances in Prostaglandin and Thromboxane Research, 3, page 113, (1978). Generally, these metabolites are beneficial but in certain disease processes or other conditions the excessive
20 production of arachidonic acid metabolites causes deleterious consequences such as inflammation, erythema, platelet aggregation, or allergic responses. The inhibition of phospholipase A2 prevents these and similar conditions.

The actual inhibition of phospholipase A2 takes place on a
25 cellular level, therefore, administration of phospholipase A2 inhibitory compounds can be by any manner that will allow for phospholipase A2 inhibition in the affected tissues or organs. The precise mechanisms of the disease processes or conditions which stimulate the arachidonic acid cascade are not clearly understood.
30 The essential prerequisite, however, is enhanced activity of the phospholipases which provide arachidonate to the series of reactions designated as the arachidonic acid cascade. One aspect of the present invention is to block the action of the phospholipases and cut off the flow of arachidonic acid into the cascade, irrespective
35 of the stimulus or stimuli which may be present. Thus, the inhibition of phospholipase A2 of this invention is suitable for treating seemingly unrelated diseases whose common element is the stimulation

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of the arachidonic acid cascade.

Hyperglycemia refers to a condition commonly found in patients suffering from mature-onset diabetes mellitus or other diseases which cause impairment of pancreatic function. Hyperglycemic patients with
5 non-insulin dependent diabetes mellitus (NIDDM) with insulin resistance exhibit elevated serum glucose levels. Failure to adequately control elevated serum glucose levels can cause myocardioischemia, stroke, peripheral vascular disease, lethargy, coma, blindness, kidney failure or death. One important means of treating these
10 patients uses oral antidiabetic agents instead of conventional treatment for hyperglycemic conditions such as restriction of carbohydrate intake or insulin injection.

INFORMATION DISCLOSURE

Some inhibitors of phospholipase A2 are known. Certain local
15 anesthetics have been shown to inhibit phospholipase A2 activity by competing with calcium ion, which appears to be necessary for phospholipase activity, see W. Vogt, Advances in Prostaglandin and Thromboxane Research, 3, p. 89 (1978) and E. Vallee et al., J. Pharm. Pharmacol., 31, pp. 588-92 (1974). R.J. Flower and G.J. Blackwell
20 have shown that certain anti-inflammatory steroids induce biosynthesis of a phospholipase A2 inhibitor which prevents prostaglandin generation, see Nature, 278, p. 456 (1979). These steroids are not direct inhibitors of phospholipase A2, but rather stimulate the synthesis of a phospholipase inhibiting factor called lipocortin,
25 lipomodulin, or macrocortin.

Some direct phospholipase A2 inhibitors are known. Indomethacin, a drug with anti-inflammatory properties, has been shown to inhibit phospholipase A2 enzymes isolated from the venoms of Russell's Viper, Crotalus adamanteus, and bee, and from pig pancreas;
30 see K.L. Kaplan, et al., Proc. Natl. Acad. Sci., 75, pp. 2955-2988 (1978). Bromphenacyl bromide has been shown to inhibit phospholipase A2 by acylating a histidine residue which is at the active site of the enzyme; see M. Roberts, et al., J. of Biol. Chem., 252, pp. 2405-2411 (1977). Mepacrine has been shown to inhibit the activity
35 of phospholipase A2 derived from perfused guinea pig lung, see R. Blackwell, et al., British J. Pharmacy, 62, p. 79-89 (1978). Some butyrophrenones are disclosed as phospholipase A2 inhibitors in U.S.

Patent 4,239,780. D. P. Wallach and V. J. R. Brown, Bioch. Pharmacol., 30, pp. 1315-24 (1981) also refer to several compounds that inhibit phospholipase A2.

U.S. Patent 3,370,070 discloses amino-substituted steroid compounds which are useful as hypocholesterolemic agents, antibacterial, anti-protozoal, and anti-algal agents. The doctoral thesis of L.J. Griggs, "Part I. Synthetic Approaches to 5- and 16-Thiaestrone. Part II. Estrone with a Diazacholesterol Side Chain," University of Michigan (1965) discloses amino-substituted steroid compounds with potential hypocholesterolemic activity. U.S. Patent 3,284,475 and P.D. Klimstra, et al., "Hypocholesterolemic Agents. VI. A- And B-Ring-Modified Azacholesterol", J. Med. Chem., 9, pp. 323-26 (1966) also disclose amino-substituted steroid compounds.

SUMMARY OF THE INVENTION

The present invention is: compounds of formula I wherein R is selected from the group consisting of $\text{CH}_2=\text{CH}-\text{CH}_2-$, $\text{HO}-\text{CH}_2\text{CH}_2\text{CH}_2-$, and CH_3 , wherein R_1 is selected from the group consisting of formulas IV-VI, wherein R_2 and R_3 are methyl or hydrogen, wherein R_4 is hydrogen or hydroxy and pharmacologically acceptable salts thereof; compounds of formula II wherein R is $(\text{CH}_3)_2\text{NCH}_2\text{CH}_2\text{CH}_2-$ or $\text{NH}_2\text{CH}_2\text{CH}_2\text{CH}_2-$ and pharmacologically acceptable salts thereof; and compounds of formula III wherein the dashed line indicates that the 2-3 bond is saturated or unsaturated and wherein R is hydrogen or methyl.

Compounds represented by formula II are useful when it is medically necessary or desirable to inhibit phospholipase A2 in a mammalian system. They are particularly useful in treating symptoms or conditions resulting from metabolites produced by the arachidonic acid cascade, hereafter PMC (phospholipase mediated conditions). PMC includes all untoward conditions or symptoms which are the result of the excessive stimulation of the arachidonic acid cascade. These conditions include allergic diseases, inflammatory conditions (including chronic inflammatory conditions such as rheumatoid arthritis), burns, and hypoxic conditions at the cellular level such as coronary infarcts or infarcts of other tissues. In infarct conditions it is desirable to block phospholipase A2 activity to prevent the destruction of the phospholipids which are integral structural components of cellular membranes.

Compounds represented by formulas I and III invention are useful as hypoglycemic agents in treating patients suffering from elevated serum glucose levels resulting from an impairment of tissue response to insulin and/or an impairment of pancreatic islet function such as non-insulin dependent diabetes mellitus (NIDDM) with insulin resistance.

DETAILED DESCRIPTION

The dosage regimen for preventing or treating phospholipase mediated conditions, PMC, by the compounds represented by formula II are selected in accordance with a variety of factors, including the type, age, weight, sex, medical condition of the mammal, severity of the PMC and the particular compound employed. An ordinarily skilled physician or veterinarian will readily determine and prescribe the effective amount of the compound to prevent or arrest the progress of the condition. Typically, the physician or veterinarian employs relatively low dosages at first and then increases the dose until a desired or maximum response is obtained. Because the diseases or conditions caused by the arachidonic acid cascade are varied, methods of administering these compounds to patients must depend on the particular PMC to be treated. Preferred patients to be treated are domesticated animals and humans, the most preferred patients are humans.

Regardless of the route of administration selected, the compounds used are formulated into pharmaceutically acceptable dosage forms by conventional methods known to the pharmaceutical art. Thus, the compounds can be administered orally in forms such as pills, capsules, solutions or suspensions; rectally or vaginally in forms such as suppositories or bougies; parenterally, either subcutaneously, intravenously, or intramuscularly using sterile injectable forms known to the pharmaceutical art. For treatment of conditions such as erythema the compounds of this invention may also be administered topically in the form of ointments, creams, gels, or the like.

Initial dosages of the compounds of this invention are from about 0.003 to 3.0 g per 70 kg mammal per 6-8 hours orally. When other forms of administration are employed, equivalent doses are administered. When dosages beyond 45 mg/kg are employed, care should be taken with each subsequent dose to monitor possible toxic effects.

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The phospholipase A2 inhibitory compounds of the present invention ameliorates the cellular damage resulting from the degradation of cell membranes by phospholipase A2 after hypoxic states such as coronary infarcts, ligation of the aorta during surgery for aortic aneurysms (resulting in kidney damage), and the like, see Zalewski, et al., Clinical Research 31, p. 227 (1983). Preferred patients to be treated are domesticated animals and humans, the most preferred patients are humans. This is a preferred use of these compounds.

The phospholipase A2 inhibitory compounds are useful in the treatment of asthma. Asthma is a lung disease in which a wide variety of stimuli can result in an asthmatic attack. These stimuli could be damp cold air or allergens in the environment. The asthmatic response is initially characterized by constriction of the bronchioles leading to increased airway resistance. This early constrictive phase is due to mast cell release of histamine and other modulators such as peptides. After the constrictive phase, a late sustained phase occurs which, in human beings, may reach a maximum in 6-8 hours. This phase is slower in onset and disappearance and is due to metabolites of the arachidonic acid cascade such as thromboxanes, prostaglandins, and leukotrienes, see "Corticosteroid Treatment in Allergic Airway Diseases, "Proceedings of a Symposium in Copenhagen Oct. 1-2, 1981 (Editors: T.H. Clark, N. Mygind, and O. Selroos, Munksgaard/Copenhagen 1982). Inhibition of phospholipase A2 at a physiologically acceptable level will prevent formation of these products in the lung thought to be responsible for the "2nd wave" of airway resistance. Preferred patients to be treated are domesticated animals and humans, the most preferred patients are humans.

For these uses, the compounds are administered in a variety of dosage forms: orally in the form of tablets, capsules, or liquids; rectally in the form of suppositories; parenterally, subcutaneously, intradermally, or intramuscularly, with intravenous administration being preferred in emergency situations, by inhalation in the form of aerosols or solutions for nebulizers; or by insufflation in the form of powder. These compounds are effectively administered to human asthma patients by oral or aerosol inhalation.

Doses in the range of about 0.01 to 50 mg per kg of body weight are used 1 to 4 times a day, the exact dose depending on the age,

weight, condition of the patient and frequency and route of administration. For the above use these compounds can be combined advantageously with other anti-asthmatic agents, such as sympathomimetics (isoproterenol, phenylephrine, ephedrine); xanthine derivatives (theophylline, aminophylline); and corticosteroids (prednisolone).

Administration by oral inhalation with conventional nebulizers or by oxygen aerosolization conveniently provides the active ingredient in dilute solution, preferably at concentrations of about 1 part of medicament to from about 100 to 200 parts of weight of total solution. Entirely conventional additives may be employed to stabilize these solutions or to provide isotonic media, for example, sodium chloride, sodium citrate, citric acid, sodium bisulfite, and the like can be employed.

A self-propelled dosage unit suitable for inhalation therapy for administering the active ingredient in aerosol form comprises the active ingredient suspended in an inert propellant such as a mixture of dichlorodifluoromethane and dichlorotetrafluoroethane together with a co-solvent such as ethanol, flavoring materials and stabilizers. Instead of a co-solvent, a dispensing agent such as oleyl alcohol can also be used. Suitable means to employ the aerosol inhalation therapy technique are described fully in U.S. Patent No. 2,868,691.

The compounds represented by formula II also control spasm and facilitate breathing in conditions such as bronchial asthma, bronchitis, bronchiectasis, pneumonia and emphysema. To treat a patient with these symptoms, the compounds would be administered to the patient in the same manner as described above. This is a preferred use of compounds represented by formula II.

These novel phospholipase A2 inhibitory compounds are also useful as anti-inflammatory agents in mammals, especially humans, and for this purpose, are administered systemically and preferably orally. For oral administration, a dose range of 0.05 to 50 mg per kg of human body weight is used to give relief from pain associated with inflammatory disorders such as rheumatoid arthritis. They are also administered intravenously in aggravated cases of inflammation, preferably in a dose range 0.01 to 100 g per kg per min until relief

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from pain is attained. When these novel compounds are administered orally, they are formulated as tablets, capsules, or as liquid preparations, with the usual pharmaceutical carriers, binders, and the like. For intravenous use, sterile isotonic solutions are preferred.

These novel phospholipase A2 inhibitory compounds are useful whenever it is desired to inhibit platelet aggregation, reduce the adhesive character of platelets, or remove or prevent the formation of thrombi in mammals. For example, these compounds are useful to prevent myocardial infarcts, to prevent post-operative thrombosis, to promote patency of vascular grafts following surgery, or to treat conditions such as atherosclerosis, arteriosclerosis, blood clotting defects due to lipemia, and other clinical conditions. Preferred patients to be treated are domesticated animals and humans, the most preferred patients are humans. For these uses, these compounds are administered intravenously, subcutaneously, intramuscularly, or in the form of sterile implants for prolonged action. For rapid response especially in emergency situations, the intravenous route of administration is preferred. Doses in the range about 0.005 to about 20 mg per kg of body weight per day are used. The exact dose depending on the age, weight, condition of the patient, and frequency and route of administration.

Phospholipase A2 Inhibitors of the present invention are useful in the treatment or prevention of conditions due to increased phospholipase activity observed after central nervous system (CNS) trauma such as brain and spinal cord injury, see, E.P. Wei, et al., J. Neurosurg., 56, pp. 695-698 (1982) and E.D. Hall and J.M. Braughler, Surgical Neurology, 18, pp. 320-327 (1982). Preferred patients to be treated are domesticated animals and humans, the most preferred patients are humans.

The compounds represented by formula II are useful to treat PMC symptoms which have already occurred in the mammal or to prevent PMC symptoms in particularly susceptible mammals. Use of these compounds before the development of PMC symptoms prevents the formation of the prostaglandins and similar products causing such conditions. Thus, phospholipase inhibitors of the present invention are used to prevent edema and erythema from sunburn by administering these compounds

prior to exposure to sunlight. These compounds are also administered to persons suffering from hayfever or similar allergies prior to exposure to the allergenic substances to which hayfever sufferers are sensitive.

- 5 It is most preferred to use the compounds of this invention in the treatment or prevention of asthma and in the treatment or prevention of cellular death resulting from hypoxic states.

10 The preferred route in most cases is to systemically administer the compounds in order to allow them to enter the mammal's bloodstream and thus be distributed throughout the mammal's system. In certain cases, where the IMC is of a localized nature (sunburn or psoriasis), topical administration is employed in order that the phospholipase A2 inhibition is confined to the afflicted area.

- 15 While conventional treatment for hyperglycemic conditions may include restriction of carbohydrate intake and insulin injection, one important means of treating hyperglycemic patients is with oral hypoglycemic agents.

20 Compounds of this invention represented by formulas I and III are useful to treat NIDDM and its complications in mammals, including human beings because these compounds lower the serum glucose levels when administered to ~~KKA^y~~ mice with spontaneous diabetes. Accordingly, a patient to be treated with certain of the novel hypoglycemic compounds of this invention is first diagnosed as a diabetic by conventional means, usually by the persistence of elevated serum
25 glucose levels, and a treatment regimen with compounds of this invention is established so that the elevation in a patient's serum glucose level is either significantly reduced or eliminated. The precise therapeutic endpoint of treatment, elimination or merely reduction in hyperglycemia, is readily determined by the attending
30 physician based upon the clinical presentation and concomitantly employed treatment. For example, certain of the novel hypoglycemic compounds of this invention may be employed to significantly reduce hyperglycemia in a patient, with a carbohydrate-restricted diet providing a further measure of control. Preferred patients to be
35 treated are domesticated animals and humans, the most preferred patients are humans.

While the novel hypoglycemic compounds of this aspect of the

invention may be administered by any convenient route; orally, subcutaneously, intravenously, intramuscularly, topically, or rectally, these compounds are most significantly and usefully employed as oral hypoglycemic agents, particularly in solid dosage form such as capsules and tablets. Alternatively, liquid oral dosage forms, such as syrups and elixirs, are alternatively employed. The solid, oral pharmaceutical compositions in accordance with the present invention are all prepared by methods known in the art for preparing other oral antidiabetic compositions. Since an individual patient response to treatment with compounds in accordance with the present invention may vary, effective dosages of the compounds of the instant invention will vary from patient to patient. Ordinarily, an oral dosage of from 0.1 to 10 mg/kg of these compounds will be adequate to significantly reduce hyperglycemia in a patient being treated. Repeated dosages, every 4-12 hr, may be required during the day to maintain the antihyperglycemic effect. Accordingly, dosages from about 0.1 mg/kg/dose to about 10 mg/kg/dose, depending upon the patient, frequency of treatment, and observed response. An attending physician may at first prescribe a relatively small amount of a novel hypoglycemic compound of this invention and later increase this dosage as necessary to achieve the desired level of control.

Novel hypoglycemic compounds of the present invention are also useful to treat and/or prevent obesity in mammals including human beings. For this purpose, the novel compounds of this invention are formulated and administered as described above for hyperglycemia.

All compounds of the present invention may be formulated into pharmaceutical compositions, employing a pharmaceutically acceptable carrier. Pharmaceutical formulations include pharmaceutical compositions suitable for oral, parenteral, vaginal, topical, and rectal use, such as tablets, powder packets, cachets, dragees, suppositories, bougies, and the like. Suitable diluents or carriers such as carbohydrates (lactose), proteins, lipids, calcium phosphate, cornstarch, stearic acid, methylcellulose and the like may be used as carriers or for coating purposes. Coconut oil, sesame oil, safflower oil, cottonseed oil, peanut oil may be used for preparing solutions or suspensions. Sweetening, coloring and flavoring agents may be added.

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In general the preferred route of administration depends on the condition being treated. For asthma, oral or aerosol inhalation is preferred. For most other conditions the preferred mode of administration is oral.

- 5 The phospholipase A2 inhibitory compounds of this invention are useful in any mammal whose metabolic system includes the phospholipase induced arachidonic acid cascade. The mammals which are preferred are generally domesticated animals and humans. Humans are the most preferred mammals to be treated.

- 10 The utility of the compounds of this invention represented by formula II is demonstrated in the following laboratory tests which determine phospholipase inhibition.

Inhibition of Rat Neutrophil Aggregation

1. Method for Thioglycolate Broth Preparation

- 15 Weigh sufficient thioglycolate medium, USP grade, to prepare a 5% w/v solution in sterile water. Heat the solution for 10 minutes on a boiling water bath. Remove and allow the solution to cool to 20-25°. Inject 10 ± 0.5 ml intraperitoneally into Sprague-Dawley rats as described below.

- 20 2. Method for Rat Peritoneal Leukocyte Collection

- Six (6) Sprague-Dawley, pathogen free, female rats (230-270 grams) are injected intraperitoneally with 10.0 ± 0.5 ml thioglycolate broth, 5% w/v 16-18 hours prior to sacrifice. After sacrifice by cervical dislocation, leukocytes accumulated in the peritoneal cavity are collected by injecting 30 ml of sterile 0.9% w/v sodium chloride intraperitoneally and vigorously massaging the abdomen to assure uniform dispersion of the cells within the carcass. Use a pasteur pipet to remove approximately 20 ml of fluid with suspended cells from a small incision through the abdominal wall. Collect the cell suspension in plastic culture tubes.

- 30 3. Washing and Resuspension of Cells for Aggregation

- Centrifuge the cell suspensions isolated above for 10 minutes at 1000 rpm (Sorvall RC-3, HG-4L rotor, 25°C). Discard the supernatant. Resuspend the cells evenly in 0.9% NaCl to original volume, centrifuge a second time for 10 minutes at 1000 rpm. Discard the supernatant. Resuspend the cells evenly in Hanks buffer.

- 35 4. Determination of Leukocyte Concentration

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Transfer 10 μ l of leukocyte suspension into a plastic cell counting cup. Add 15.0 ml of ISOTON- diluent for cell counting. Determine the cell count with a Model ZBI Coulter Counter or equivalent.

5 5. Neutrophil Aggregation

A. Add 0.5 ml of rat leukocyte (neutrophil) suspension to each channel of a Payton dual channel aggregometer. Cuvettes, 45 mm x 4 mm i.d., are used. Cell suspensions at 37°C are stirred (400 rpm).

10 B. Add 5 μ l test compound (.01 M in absolute ethanol) to cell and evaporate to dryness under nitrogen. Add 0.5 ml cell suspension (37°C, 400 rpm). Incubate for 2 minutes, then add 1 μ l of the agonist, 10^{-4} M FMLP.

C. Record the aggregation trace (% transmitted light) on a potentiometric recorder.

15 Inhibition of Hog Pancreas PLA₂

1. Enzymes

Both soybean lipoxidase and hog pancreas phospholipase A₂ are obtained commercially from Sigma. The soybean lipoxidase is dissolved at a concentration of 5 mg/ml in 0.033 M ammonium chloride buffer pH 8.5 with 1×10^{-4} M Ca⁺⁺. The hog pancreas enzyme is added at the rate of approximately 350 units per ml of final mixture. Thus, 0.025 ml is equivalent to 9 units of phospholipase and 0.125 mg of lipoxidase.

2. Substrate

25 The substrate is phosphatidyl choline. The material has a fatty acid composition upon saponification, of 2% of 16:0, 1% of 18:0, 3% of 18:1, 18% of 18:2, and 12% of 18:3 fatty acids with the largest fraction being linoleic acid. The estimated molecular weight is 780.

30 78 mg of this substrate is put in a 10 ml volumetric flask containing 100 mg of deoxycholic acid. A "pill" magnetic stirrer is added along with 7-8 ml of water, and the whole stirred rapidly until all the lecithin is dissolved. The "pill" is then removed and the flask contents are made up to 10 ml with water.

35 3. Procedure

To three oxygraph cells equipped with magnetic stirrers is added 2.5 ml of 0.033 M ammonium chloride buffer, pH 8.5, containing 1×10^{-4} M

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Ca^{++} . This is followed by 0.1 ml of the inhibitors made up at an initial concentration of 0.01 M in methanol. Where controls are run, 0.1 ml of methanol is added to each cell. The cells are then put in the oxygraph apparatus and the contents are stirred briefly. 0.025 ml of the enzyme mixture is then added and the electrodes are inserted in each cell, care being taken to exclude all air bubbles. With the stirrer and water bath pump on, the contents of each cell are stirred for 2.5 minutes at 37.5°C. The reaction is initiated by adding to the cells 0.05 ml of 0.01 M lecithin substrate. The reaction is monitored by continuous measurement of rates of oxygen depletion from the medium as a consequence of unsaturated fatty acids (linoleic acid) being released from esterified form by the phospholipase. These fatty acids immediately become substrates for the soybean lipoxidase which forms the 15-hydroxy acids, with consequent oxygen utilization.

The initial rates of oxygen consumption are recorded using a Sargeant-Welch Recorder set at 5 mV full scale. The "air" setting and medium chart speed are used. The slopes of oxygen consumption are then determined in triplicate, and these are compared with the methanol controls to determine the degree of inhibition. If complete inhibition is seen at the first concentration, appropriate dilutions are made to bring the inhibition percentages down to at least 3 concentrations of inhibitor where partial inhibition is observed. The I_{50} can then be calculated for that particular inhibitor, using linear regression slopes. All compounds for which I_{50} value is shown are tested for inhibitory activity on the soybean lipoxidase. None inhibit at the test concentration.

The utility of the compounds of this invention represented by formulas I and III is demonstrated in the following laboratory tests which determine serum glucose levels in mice.

Testing For Blood Glucose Lowering In The KKA^y House

1. General

All KKA^y mice used for screening are produced and selected by methods previously outlined; T. Fujita et al., Diabetes, 32, 804-10 (1983). Groups of 6 animals each are employed.

2. Screening Procedure

Pre-treatment nonfasting blood glucose (NFBG) samples are

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- measured 5 days prior to the start of a screening run by previously described methodologies. These blood sugar values are used to place animals into groups with equal mean blood glucose concentrations and to eliminate any mice with a NFBG value <250 mg/dl. On day 0, compounds chosen to be run are incorporated into ground mouse chow (Purina 5015). Compounds are included at a rate of 1 mg/gram of chow. Generally, 300 g of drugs containing diet is prepared for each group. Mice receiving ground chow only are the negative control. Each screening run also uses ciglitazone (T. Fujita, et al., supra) as a positive control (0.5 to 1.0 mg/gram chow).

Initial body and food weights are taken on day 1. Food is placed in a crock which contains an adequate amount to last for the length of the study. In order to acclimate the mice from pelleted mouse chow to ground mouse chow, they are fed the ground chow for 9 days prior to use in the screen. On day 4 of treatment, a NFBG sample is again measured, as well as food and body weights. Food consumption measurements are used to determine an average mg/kg dose the mice received over the -testing period, and to evaluate the compound's effect on food consumption.

3. Acceptance of a Screening Run and Determination of Activity

Acceptance and activity are determined by the following criteria:

A. Negative Control

This group must not show a significant change ($p < .05$) from pre- to post-treatment. If there is a significant decrease in blood sugar the run is not valid.

B. Positive Control

This group must show a significant depression in blood sugar mean levels from pre- to post-treatment. A lack of activity in this group would also invalidate the run.

C. Negative Control vs. Positive Control

This contrast must be significant. It is a further assurance that both control groups performed as expected.

D. Compound

A compound's activity is based on several criteria:

(1) A significant decrease in blood sugar mean levels from pre- to post-treatment.

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(2) Negative control vs. compound: This contrast allows one to determine if these groups are dissimilar, which is required for the compound to be considered active.

General syntheses of compounds similar to those of the present invention are set forth in PCT application PCT/US86/02116, filed 7 October 1986, which is incorporated herein by reference.

A bond indicated as "-" includes both the α and β configurations.

Compounds that have been found to have a phospholipase A_2 inhibitory or antidiabetic effect as determined by at least one of the above assays are indicated in the Examples and Preparations which follow by the notation "PLA₂" and/or "diabetes" respectively.

EXAMPLES

Preparation 1 17 β -t-Butyldimethylsilyloxy-19-nor-androstan-3-one

A 250 ml, 3-necked flask, equipped with a magnetic stirrer, was flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 10 g of 17 β -hydroxy-5 α -estran-3-one dissolved in 150 ml of dimethylformamide. The solution was treated with 3.7 g of imidazole and the solution was cooled to 0°C. The solution was then treated with 6.5 g of t-butyldimethylsilylchloride and allowed to stir at room temperature for 48 hours. The reaction mixture was diluted with water and extracted twice with hexane/ethyl acetate (9:1). The combined organic layers were washed with brine, dried over anhydrous magnesium sulfate and concentrated in vacuo.

The crude product (14.6 g) was flash chromatographed on 90 g of 230-400 mesh silica gel. The column was packed and eluted with (91:9) hexane/ethyl acetate with (no column volume eluted) 30 ml fractions collected. Based on their TLC homogeneity, fractions 4-13 were combined affording 14 g of the title compound, m.p. 103-104°C.

Preparation 2 17 β -t-Butyldimethylsilyl-19-nor-androstan-3,17-diol

A 2000 ml, 3-necked flask, equipped with a magnetic stirrer, was flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 14 g of the title compound from Preparation 1 dissolved in 850 ml of methanol/methylene chloride (15:2). The solution was treated with 7 g of sodium borohydride in small portions. The

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reaction mixture was allowed to stir for 15 minutes. The reaction mixture was quenched with 2 M NaHSO₄, diluted with water and extracted with ethyl acetate. The organic layer was washed with water, washed with brine, dried over anhydrous magnesium sulfate, and concentrated in vacuo.

The crude product (15.3 g) was chromatographed on 1000 g of 230-400 mesh silica gel. The column was packed and eluted with (98:2) methylene chloride/acetone. An initial fraction of 1200 ml was collected followed by 17 ml fractions. Based on their TLC homogeneity, fractions 280-430 were combined affording 10.9 g (77% of theory) of the title compound (β -isomer), m.p. 135-138°C.

Preparation 3 3-[[$(3\beta,5\alpha)$ -17-[(*t*-Butyldimethylsilyl)oxy]estrane-3-yl]oxy]propanenitrile

A 250 ml, 3-necked flask, equipped with a magnetic stirrer, was flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 8.0 g of 3β alcohol steroid from Preparation 2 dissolved in 150 ml of benzene. The solution was treated with 2.2 ml of freshly distilled acrylonitrile followed by 0.37 ml of Triton B and the reaction mixture was stirred at room temperature for 72 hours. The reaction mixture was washed with dilute HCl, washed with water, dried over anhydrous magnesium sulfate and concentrated in vacuo.

The crude product (10.9 g) was chromatographed on 1070 g of 230-400 mesh silica gel. The column was packed and eluted with (83:17) hexane/ethyl acetate. An initial fraction of 1500 ml was collected followed by 18 ml fractions. Based on their TLC homogeneity, fractions 81-110 were combined affording 7.0 g of the title compound, NMR (CDCl₃, TMS): 6 3.8-3.4(m), 3.4-3.0(br), 2.7-2.45(t), 2.2-0.4(m), 0.9(s) and 0.75 ppm(s).

Preparation 4 3-[[$(3\beta,5\alpha)$ -17-[hydroxy]estrane-3-yl]oxyl]propanenitrile

A 500 ml, 3-necked flask, equipped with a magnetic stirrer, was flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 4.4 g (9.9 mmol) of 3β -cyano ether from Preparation 3 dissolved in 500 ml of methylene chloride and 200 ml of methanol. The solution was treated with 32 ml (102 mmol) of a 3.2 M HCl in methanol solution. The reaction mixture was allowed to stir at room temperature for 30 min. The reaction mixture was diluted with water

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and extracted with ethyl acetate. The organic layer was washed again with water, washed with brine, dried over anhydrous magnesium sulfate and concentrated in vacuo.

The crude product (3.73 g) was flash chromatographed on 90 g of 230-400 mesh silica gel. The column was packed and eluted with (98:2) methylene chloride/acetone (no column volume eluted) with 25 ml fractions collected. Based on their TLC homogeneity, fractions 8-25 were combined affording 3.1 g of the title compound. NMR (CDCl₃, TMS): 6 3.75-3.4 (m), 3.4-3.0 (br, 2.6-2.3 (t), 2.15-0.75 (m) and 0.75 ppm (s).

Preparation 5 3[[[(3 β ,5 α)-17-estran-one]oxy]propanenitrile

A 50 ml, 3-necked flask, equipped with a magnetic stirrer, was flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 1.0 g of 3 β -17-hydroxy steroid from Preparation 4 dissolved in 20 ml of acetone and the solution was cooled to 0°C. The solution was treated with 1.0 ml of Jones reagent reaction and the mixture was stirred for 15 minutes. The reaction mixture was quenched with 5 ml of 2-propanol. The reaction mixture was diluted with water and extracted with ethyl acetate. The organic layer was washed with water, washed with brine, dried over anhydrous magnesium sulfate, and concentrated in vacuo, to give 1.1 g of the title compound. NMR (CDCl₃, TMS): 6 3.8-3.55 (t, 2H), 3.5-3.1 (br, 1H), 2.8-2.45 (t, 2H), 2.45-0.95 (m) and 0.9 ppm (s, 3H).

Preparation 6 N-[(3 β ,5 α)-3-((3-propanenitrile)oxy)estran-17-yl]-1,3-propanediamine

A 50 ml, 2-necked flask, equipped with a magnetic stirrer, was flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 0.36 ml of 1,3-diaminopropane dissolved in 10 ml of methanol. The pH of the solution was adjusted to 6.0 with glacial acetic acid. The solution was then treated with 282 mg of 3 β -steroid from Preparation 5 and followed by 75 mg of sodium cyanoborohydride. The reaction mixture was refluxed for 18 hours. The reaction mixture was basified with concentrated ammonium hydroxide and then concentrated in vacuo.

The crude product (3.29 g) was chromatographed on 100 g of 230-400 mesh silica gel. The column was packed and eluted with (92.3:7.0:0.7) chloroform/methanol/ammonia. An initial fraction of 150 ml

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was collected, followed by 6 ml fractions. Based on their TLC homogeneity, fractions 45-180 were combined affording 250 mg of the title compound. NMR (CDCl₃, TMS) δ 3.75-3.55 (t, 2H), 3.4-3.0 (br, 1H), 2.8-0.75 (m) and 0.65 ppm (s, 3H).

5 Preparation 7 N-(3 β ,5 α)-3-(3-aminopropoxy)estrane-17-ol

A 250 ml, 3-necked flask, equipped with a magnetic stirrer, was flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 105 mg of lithium aluminum hydride slurried in 30 ml of diethyl ether. The solution was treated with 230 mg of 3 β -ether-17-
10 hydroxy steroid from Preparation 4 dissolved in 50 ml of diethyl ether and the reaction mixture was refluxed for 2 hours. The reaction mixture was quenched with 0.21 ml of water followed by 0.17 ml of a 10% sodium hydroxide solution and the reaction mixture was allowed to stir at room temperature for 18 hours. The reaction
15 mixture was filtered and the solids washed several times with hot chloroform. The filtrate was concentrated in vacuo.

The crude product (350 mg) was flash chromatographed on 90 g of 230-400 mesh silica gel. The column was packed and eluted with (92.3:7.0:0.7) chloroform/methanol/ammonia. An initial 5-15 ml
20 fractions were collected followed by 30 ml fractions. Based on their TLC homogeneity, fractions 8-14 were combined affording 175 mg of the title compound. NMR (CDCl₃, TMS): δ 3.56-3.51 (t, 2H), 3.25-3.1 (br, 1H), 2.8-2.77 (t, 2H), 2.05-0.92 (m and 0.74 ppm (s, 3H).

Preparation 8 N-(3 β ,5 α)-3-(3-aminopropoxy)estrane-17-one

25 A 25 ml, 2-necked flask, equipped with a magnetic stirrer, was flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 170 ml of 3 β -aminoether-17-hydroxy steroid from Preparation 7 dissolved in 5 ml of acetone and then cooled to 0°C. The solution was treated with 27 μ l (0.51 mmol) of concentrated sulfuric
30 acid followed by 0.31 ml of Jones reagent and was allowed to stir for 1 hour. The reaction mixture was quenched with 2 ml of 2-propanol followed by 0.6 g of sodium citrate dihydrate and a small piece of amalgamated zinc (Org. Syn. Coll., Vol. IV, p. 696 (1963). The reaction mixture was allowed to stir for 30 minutes at room tempera-
35 ture. The reaction mixture was basified with 3 M KOH (pH 10) and the aqueous layer saturated with sodium chloride. The aqueous layer was extracted 5 times with methylene chloride, 2 times with chloroform

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and once with diethyl ether. The aqueous layer was then stirred vigorously with methylene chloride. The combined organic extracts were dried over anhydrous magnesium sulfate and concentrated in vacuo.

- 5 The crude product (85 mg) was chromatographed on 8 g of 230-400 mesh silica gel. The column was packed and eluted with (92.3 : 7.0 : 0.7) chloroform/methanol/ammonia. An initial fraction of 5 ml was collected followed by 0.8 ml fractions. Based on their TLC homogeneity, fractions 23-40 were combined affording 45 mg of the title
10 compound. Infrared: λ_{max} (chloroform solution) 2950, 2850 and 1740 cm^{-1} . (PLA2)

Preparation 9 N-[(3 β ,5 α)-3-[3-(dimethylamino)propoxy]estrane-17-one]

- A 10 ml, 2-necked flask, equipped with a magnetic stirrer, was
15 flame dried and then cooled in a nitrogen atmosphere. The flask was charged with 44 mg of 3 β -aminoether-17-keto-steroid from Preparation 8 dissolved in 1 ml of dioxane. The solution was treated with 1.3 ml of a 1.0 M monosodium phosphorous acid solution followed by 98 μl of 37% formaldehyde. The reaction mixture was then heated at 60°C for 2
20 hours. The reaction mixture was diluted with methylene chloride and water. The aqueous layer was basified with 3 M KOH to a pH of 11. The organic layer was separated and the aqueous layer was re-extracted 2 times with methylene chloride. The combined organic layers were washed with brine, dried over anhydrous magnesium sulfate and
25 concentrated in vacuo.

The crude product (36 mg) was not chromatographed. Based on TLC, crude product was determined to be relatively pure title compound. NMR (CDCl_3 , TMS): δ 3.6-3.4 (m), 3.3-2.8 (m), 2.7 (s), 2.6-0.4 (m) and 0.8 ppm (s).

- 30 Example 1 3-Methoxy-17 β -[(2-thienylmethyl)amino]-estra-1,3,5-(10)-triene

- A solution of 7.96 g of 2-thiophenemethylamine in 50 ml of MeOH and 150 ml of THF was acidified with 6 ml (6.29 g) of glacial acetic acid. Then 10 g of estrone methyl ether was added. The mixture was
35 heated until a solution was obtained and then stirred at ambient temperature for 1 hour. Sodium cyanoborohydride (2.18 g) was added. The resulting solution was stirred for 5 hours. An additional 2.18 g

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of sodium cyanoborohydride was added. The stirring was continued for 17 hours. The solvent was evaporated. The residue was treated with 200 ml of H₂O and basified with a 50% NaOH solution. The mixture was extracted with CH₂Cl₂ (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO₄. Evaporation of the solvent left 17.54 g of oil. The oil was chromatographed on a 1100 g column of silica gel. The column was eluted with 7.5% MeOH-CH₂Cl₂ and 200 ml fractions were collected. The fractions were assayed by silica gel TLC (1 x 4") (5% MeOH-CH₂Cl₂). Fractions 14-22 were combined and crystallized from CH₂Cl₂-hexane affording 9.0 g of the title compound as clusters of pale yellow needles, m.p. 85.5-87°. (diabetes)

Example 2 3-Allyloxy-17β-(((3-trifluoromethyl)phenylmethyl)-amino-estra-1,3,5(10)-triene Fumarate

A solution of 4 g of 3-(trifluoromethyl)benzylamine in 25 ml of MeOH and 75 ml of THF was acidified with 3 ml (3.15 g) of acetic acid. Then 5 g of 3-allyloxyestrone was added. After a solution was obtained, 1.2 g of sodium cyanoborohydride was added. The resulting solution was stirred for 3 hours. An additional 1.2 g of NaCNBH₃ was added. The stirring was continued for 66 hours. The solvent was evaporated. The residue was treated with 200 ml of H₂O basified with 50% NaOH solution, and extracted with CH₂Cl₂ (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO₄. Evaporation of the solvent left a pale yellow oil. The oil was chromatographed on a 700 g column of silica gel. The column was eluted with 10% acetone-CH₂Cl₂ and 200 ml fractions were collected. The fractions were assayed by silica gel TLC (1 x 4") (10% acetone-CH₂Cl₂). Fractions 9-18 were combined giving 6.83 g of a pale yellow oil. A solution of the 6.83 g of oil in 100 ml of acetone was added to a solution of 1.68 g (14.47 μmoles) of fumaric acid in 30 ml of EtOH. The solution was concentrated and then hexane was added. Cooling gave 7.56 of the title compound as a white solid, m.p. 171-174°. (diabetes)

Example 3 3-(3-Hydroxypropoxy)-17β-(((3-trifluoromethyl)phenylmethyl)amino-estra-1,3,5(10)-triene Fumarate (1:1)

A solution of 2.5 g of 3-(trifluoromethyl)benzylamine in 25 ml of MeOH and 75 ml of THF was acidified with 3 ml (3.15 g) of acetic

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acid. Then 2.37 g of 3-(3-hydroxypropoxy)-estra-1,3,5(10)-trien-17-one was added. After a solution was obtained, 1.2 g of sodium cyanoborohydride was added. The resulting solution was stirred for 4 hours. An additional 1.2 g of NaCNBH_3 was added. The stirring was continued for 18.5 hours. The solvent was evaporated. The residue was treated with 200 ml of H_2O basified with 50% NaOH solution, and extracted in the CH_2Cl_2 (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO_4 . Evaporation of the solvent left a pale yellow oil. The oil was chromatographed on a 400 g column of silica gel. The column was eluted with 25% acetone- CH_2Cl_2 and 100 ml fractions were collected. The fractions were assayed by silica gel TLC (1 x 4") (25% acetone- CH_2Cl_2). Fractions 16-23 were combined giving 2.94 g of a pale yellow oil. A solution of the 2.94 g (6.03 mmoles) of oil in 75 ml of acetone was added to a solution of 0.7 g (6.03 mmoles) of fumaric acid in 20 ml of EtOH. The solution obtained was concentrated and the hexane was added. Cooling gave 2.54 g (70%) of the title compound as a white solid, m.p. 117-121° (diabetes).

Example 4 3-Methoxy-7 α -methyl-17 β -[[(3-trifluoromethyl)phenyl-methyl]amino]-estra-1,3,5(10)-triene Fumarate Hydrate

A solution of 3.32 g of 3-(trifluoromethyl)benzylamine in 25 ml of MeOH and 75 ml of THF was acidified with 3 ml (3.15 g) of acetic acid. Then 2.83 g of 7 α -methylestrone methyl ether was added. After a solution was obtained, 1.2 g of sodium cyanoborohydride was added. The resulting solution was stirred for 6 hours. An additional 1.2 g of NaCNBH_3 was added. The stirring was continued for 17 hours. The solvent was evaporated. The residue was treated with 200 ml of H_2O , basified with 50% NaOH solution, and extracted with CH_2Cl_2 (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO_4 . Evaporation of the solvent left a pink oil. The oil was chromatographed on a 400 g column of silica gel. The column was eluted with 10% acetone- CH_2Cl_2 and 100 ml fractions were collected. The fractions were assayed by silica gel TLC (1 x 4") (10% acetone- CH_2Cl_2). Fractions 10-19 were combined giving 3.93 g of pale yellow oil. A solution of the 3.93 g of oil in 75 ml of acetone was added to a solution of 0.5 g (4.31 mmoles) of fumaric acid in 15 ml of EtOH. The mixture was reduced in volume to 40 ml. Then 100 ml of

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hexane was added. Cooling gave 2.79 g of the title compound as a white solid, m.p. 189-190°. (diabetes)

Example 5 3-Methoxy-11 β -hydroxy-17 β -(((3-trifluoromethyl)phenyl-methyl)amino)-estra-1,3,5(10)-triene

- 5 A solution of 2.47 g of 3-(trifluoromethyl)benzylamine in 25 ml of MeOH and 75 ml of THF was acidified with 3 ml (3.15 g) of acetic acid. Then 2.12 g (7.06 μ moles) of 11 β -hydroxyestrone methyl ether was added. After a solution was obtained, 1.2 g of sodium cyanoborohydride was added. The resulting solution was stirred for 6 hours.
- 10 An additional 1.2 g of NaCNBH₃ was added. The stirring was continued for 18 hours. The solvent was evaporated. The residue was treated with 200 ml of H₂O, basified with 50% NaOH solution, and extracted with CH₂Cl₂ (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO₄. Evaporation of the solvent left an
- 15 oil. The oil was chromatographed on a 400 g column of silica gel. The column was eluted with 10% MeOH-CH₂Cl₂ and 100 ml fractions were collected. The fractions were assayed by silica gel TLC (1 x 4") (10% MeOH-CH₂Cl₂). Fraction 11 was crystallized from CH₂Cl₂-hexane giving 0.39 g of white needles. The 0.39 g was combined with fraction 12
- 20 and crystallized from CH₂Cl₂-hexane giving 1.06 g of the title compound as white needles, m.p. 122-123°. (diabetes)

Example 6 N-Methyl-17 β -[2-(4-aminosulfonylphenyl)ethyl]amino]-5 α -androstane

- 25 To a stirred solution of 3.45 g (7.52 μ moles) of crude 17 β -[2-(4-aminosulfonylphenyl)ethyl]amino]-5 α -androstane in 175 ml of THF and 125 ml of acetonitrile was added 3 ml (40 μ moles) of 37% formaldehyde solution and 1 g (15.91 μ moles) of sodium cyanoborohydride followed by 1 ml (1.05 g, 17.47 μ moles) of acetic acid. The mixture was stirred for 24 hours. The solvent was evaporated. The residue
- 30 was treated with 200 ml of H₂O, basified with 50% NaOH solution, and extracted with CH₂Cl₂ (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO₄. Evaporation of the solvent left 0.66 g of oil. The combined aqueous phases, which were milky, were acidified with acetic acid. The mixture was extracted
- 35 with CH₂Cl₂ (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO₄. Evaporation of the solvent left 3.3 g of solid. The 3.3 g appeared to be the acetate salt. A

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mixture of the 3.3 g, 100 ml of CH_2Cl_2 , and 150 ml of 10% NaHCO_3 solution was stirred for 17 hours. The layers were separated. The CH_2Cl_2 layer was dried over MgSO_4 . Evaporation of the solvent left 2.5 g of foamy solid. The aqueous layer was extracted with 100 ml of CH_2Cl_2 . The extract was dried over MgSO_4 . Evaporation of the solvent left 0.05 g of oil which was discarded. The 0.66 g and the 2.58 g were combined, dissolved in 50 ml of 1:1 acetone- CH_2Cl_2 , and applied to a 400 g column of silica gel (packed in 1:1 acetone- CH_2Cl_2). The column was eluted with 1:1 acetone- CH_2Cl_2 and 100 ml fractions were collected. Fraction 13 was crystallized from CH_2Cl_2 -Skelly B giving 0.39 g of solid. The 0.39 g was recrystallized from acetone-Skelly B giving 0.16 g of solid. The 0.16 g was combined with fractions 14-22 and crystallized from CH_2Cl_2 -hexane (cooled at room temperature overnight) giving a white solid. The solid was dried in a vacuum oven at 55° for 21 hours giving 1.46 g of the title compound, m.p. 168-172° (diabetes)

Example 7 2-Methyl-3-methoxy-17 β -[2-(4-aminosulfonylphenyl)-ethylamino]-estra-1,3,5(10)-triene

A mixture of 2.54 g (8.51 μmoles) of 2-methyl-3-methoxyestra-1,3,5(10)-trien-17-one, 3.41 g (17.03 μmoles) of 4-(2-aminoethyl)benzenesulfonamide (Interchem. Corp), and 180 ml of MeOH was heated until a solution was obtained. Then 0.6 g (9.55 μmoles) of sodium cyanoborohydride was added. The resulting solution was stirred for 3 hours. Then 1 ml (1.05 g, 17.47 μmoles) of acetic acid was added. The mixture was stirred and refluxed for 44 hours. The solvent was evaporated. The residue was treated with a solution of 8 g of NaHCO_3 in 200 ml of H_2O and extracted with CH_2Cl_2 (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO_4 . Evaporation of the solvent left 4.03 g of solid. The solid was dissolved in 50 ml of 10% MeOH- CH_2Cl_2 and applied to a 400 g column of silica gel (packed in 10% MeOH- CH_2Cl_2). The column was eluted with 10% MeOH- CH_2Cl_2 and 100 ml fractions were collected. The fractions contained some solid, m.p. ca 230, which was insoluble in CH_2Cl_2 and H_2O but was soluble in MeOH. The solid was saved. Fractions 15-23 were combined in CH_2Cl_2 and filtered to remove insoluble material. The filtrate was concentrated and hexane was added. Cooling gave 2.80 g of white solid. The 2.80 g was recryst-

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tallized from acetone-hexane giving a white solid. The solid was dried in a vacuum oven at 54° for 17 hours giving 2.60 g of the title compound, m.p. 170-173. (diabetes)

Example 8 17 β -[2-(4-Aminosulfonylphenyl)ethyl]amino]-5 α -androst-2-ene

A mixture of 4.32 g (15.86 mmoles) of 5 α -androst-2-en-17-one, 4.76 g (23.17 mmoles) of 4-(2-aminoethyl)benzenesulfonamide (Interchem Corp.), and 180 ml of MeOH was heated until a solution was obtained. Then 1 g (15.91 mmoles) of sodium cyanoborohydride was added followed by 1.5 ml (1.57 g, 26.2 mmoles) of acetic acid. The resulting solution was stirred and refluxed for 42 hours. The solvent was evaporated. The residue was treated with a solution of 5 g of NaHCO₃ in 200 ml of H₂O and extracted with CH₂Cl₂ (3 x 100 ml). The combined extracts were washed with 50 ml of brine and dried over MgSO₄. Evaporation of the solvent left 6.67 of solid. The solid was dissolved in 50 ml of 10% MeOH-CH₂Cl₂ and applied to a 400 g column of silica gel (packed in 10% MeOH-CH₂Cl₂). The column was eluted with 10% MeOH-CH₂Cl₂ and 100 ml fractions were collected. Fractions 15-25 were combined and dissolved in MeOH. Triethylamine (5 ml) was added followed by H₂O. The mixture was cooled in an ice bath. The somewhat gelatinous solid which separated was collected by filtration, washed with H₂O, and dried in a vacuum oven at 55° for 16 hours giving 6.1 g. The 6.1 g was crystallized from acetone-hexane giving a white solid. The solid was crushed and then dried in a vacuum oven at 54° for 65 hours giving 4.97 of the title compound, a.p. 158-159.5°. (diabetes)

Example 9 N-[(3 β ,5 α)-3-(3-aminopropoxy)estrane-17-yl]-1,3-propanediamine

A 500 ml Parr flask was charged with 195 mg of 38-ether-17-aminopropyl steroid N-[(3 β ,5 α)-3-((3-propanenitrile)oxy)estrane-17-yl]-1,3-propanediamine dissolved in 25 ml of a 2.5 M NH₃ in ethanol solution. The solution was treated with 100 mg of 5% rhodium on alumina and the reaction mixture was placed on the Parr hydrogenation apparatus maintained at 50 psi for 4 hrs. The reaction mixture was filtered through celite and the solids were washed with 100 ml of ethanol. The filtrate was concentrated in vacuo.

The crude product (176 mg) was chromatographed on 60 g of 230-

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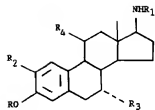
400 mesh silica gel. The column was packed and eluted with (90.0: -
9.1:0.9) chloroform/methanol/ammonia. An initial fraction of 150 ml
was collected followed by 6 ml fractions. Based on their TLC
homogeneity, fractions 160-290 were combined giving 117 mg of the
5 title compound. NMR (CDCl₃, TMS): δ 3.6-3.4 (t), 3.35-2.9 (br),
2.85-2.2 (m), 2.2-0.75 (m) and 0.65 ppm (s).

Example 10 N-[(3 β ,5 α)-3-[3-(dimethylamino)propoxy]estrane-17-yl]-
1,3-diaminopropane

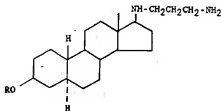
A 10 ml, 2-necked flask, equipped with a magnetic stirrer, was
10 flame dried and then cooled in a nitrogen atmosphere. The flask was
charged with 41 μ l (0.5 mmol) of 1,3-diaminopropane dissolved in 1.5
ml of methanol. The pH of the solution was adjusted to 6.0 with
glacial acetic acid. The solution was then treated with 35 ml of 3 β -
dimethylaminoether-17-keto-steroid from Preparation 9 followed by 9
15 mg of sodium cyanoborohydride and the reaction mixture was refluxed
for 5 hours. The reaction mixture was concentrated in vacuo.

The crude product (135 mg) was chromatographed on 8 g of 230-400
mesh silica gel. The column was packed and eluted with (85.7: -
12.9:1.4) chloroform/methanol/ammonia. An initial fraction of 5 ml
20 was collected, followed by 0.9 ml fractions. Based on their TLC
homogeneity, fractions 35-54 were combined affording 14 mg of the
title compound. NMR (CDCl₃, TMS): δ 3.6-3.45 (t, 2H), 3.3-3.1 (br,
1H), 2.8-2.65 (m), 2.6-2.25 (m), 2.4-2.3 (m), 2.2 (s), 2.10.8 (m) and
0.7 ppm (s, 3H). (PLA2)

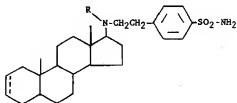
- 25 -

FORMULAS

I



II



III

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FORMULAS (continued)

IV



V

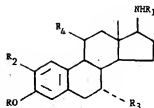


VI

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CLAIMS

1. A compound of the formula



wherein R is selected from the group consisting of

- a. $\text{CH}_2=\text{CH}-\text{CH}_2-$,
 b. $\text{HO}-\text{CH}_2\text{CH}_2\text{CH}_2-$, and
 c. CH_3 ; and

wherein R₁ is selected from the group consisting of

- a. ,
 b. , and
 c. ;

wherein R₂ and R₃ are methyl or hydrogen;
 wherein R₄ is hydrogen or -OH;
 and pharmacologically acceptable salts thereof.

2. A compound according to claim 1 selected from the group consisting of

- a. 3-methoxy-17β-[(2-thienylmethyl)-amino]-estra-1,3,5-(10)-

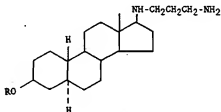
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triene,

- b. 3-Allyloxy-17 β -[((3-trifluoromethyl)-phenylmethyl)amino-estra-1,3,5(10)-triene fumarate,
- c. 3-(3-hydroxypropoxy)-17 β -[((3-trifluoromethyl)phenylmeth-
yl)amino-estra-1,3,5(10)-triene fumarate,
- d. 3-Methoxy-7 α -methyl-17 β -[((3-trifluoromethyl)phenylmeth-
yl)amino-estra-1,3,5(10)-triene fumarate hydrate,
- e. 3-Methoxy-11 β -hydroxy-17 β -[((3-trifluoromethyl)-phenyl-
methyl)amino-estra-1,3,5(10)-triene, and
- f. 2-methyl-3-methoxy-17 β -(2-(4-amino-sulfonylphenyl)eth-
yl)amino]estra-1,3,5(10)-triene,

and pharmacologically acceptable salts thereof.

3. A compound of the formula



wherein R is

- a. $(\text{CH}_3)_2\text{NCH}_2\text{CH}_2\text{CH}_2-$, or
- b. $\text{NH}_2\text{CH}_2\text{CH}_2\text{CH}_2-$;

and pharmacologically acceptable salts thereof.

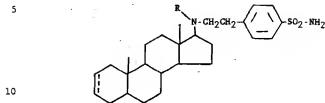
4. A compound according to claim 3 selected from

- a. N-[(3 β ,5 α)-3-(3-aminopropoxy)-estrane-17-yl]-1,3-propane-
diamine, and
- b. N-[(3 β ,5 α)-3-[(dimethylamino)-propoxy]estrane-17-yl]-1,3-
diaminopropane,

and pharmacologically acceptable salts thereof.

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5. - A compound of the formula



III

15 wherein the dashed line indicates that the 2-3 bond is saturated or unsaturated and,
wherein R is hydrogen or methyl.

6. A compound according to claim 5 selected from
- 20 a. N-methyl-17 β -[2-(4-aminosulfonylphenyl)-ethyl]amino]-5 α -androstane, and
- b. 17 β -[2-(4-aminosulfonylphenyl)ethyl]-amino]-5 α -androst-2-ene,
and pharmacologically acceptable salts thereof

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US 88/00978

I. CLASSIFICATION OF SUBJECT MATTER (In several classification symbols solely, indicate all) ¹	
According to International Patent Classification (IPC) or to both National Classification and IPC	
IPC ⁴ : C 07 J 41/00; A 61 K 31/565	
II. FIELDS SEARCHED	
Minimum Documentation Searched ⁷	
Classification System	Classification Symbols
IPC ⁴	C 07 J 41/00
Documentation Searched other than Minimum Documentation to the extent that such Documents are included in the Fields Searched ⁸	

III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹

Category ¹⁰	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
X	US, A, 3107255 (R.E. COUNSELL) 15 October 1963 see the whole document	3, 4
P, X	WO, A, 87/02367 (THE UPJOHN COMPANY) 23 April 1987 see claims	1-5

* Special categories of cited documents: ¹⁴

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

- "X" document of particular relevance; the claimed invention cannot be considered novel or distinct be considered to involve an inventive step
- "Y" document of particular relevance; the claimed invention cannot be considered novel or distinct when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "A" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search
11th July 1988

Date of Mailing of the International Search Report
26 JUL 1988

International Searching Authority

EUROPEAN PATENT OFFICE

Signature of Authorized Officer

M. VAN MOL

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SA 21935

For more details about this annex : see *Official Journal of the European Patent Office*, No. 12/82